

ECM remodelling in IBD: innocent bystander or partner in crime?

The emerging role of extracellular molecular events in sustaining intestinal inflammation

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INTRODUCTION

IBD, which primarily includes UC and Crohn's disease (CD), is a progressive, chronic and relapsing condition. This debilitating disease is steadily becoming a worldwide medical concern, with increasing prevalence and incidence in both industrialised and developing countries.¹ While the exact aetiology of the disease remains unknown, genetic predisposition and various environmental and immunological causes have been identified as contributing factors.² Generally, IBD is characterised by a dysregulated excessive immune response and tissue damage in the GI tract.^{3,4} This aberrant and sustained immune response is thought to be mainly facilitated by defects in the function of the intestinal epithelial barrier and in the regulation of mucosal immunity.⁵ Yet, tissue damage associated with IBD is commonly considered solely a downstream effect and not a contributing factor. This view has led to a concentrated focus on the development of IBD treatments that target inflammatory pathways, but all, thus far, have exhibited limited efficacy. In contrast, none of the IBD drugs released to date were designed to specifically target the tissue-destructive processes associated with the disease.

When it comes to basic and clinical research of IBD-associated tissue destruction, most groups have directed their efforts to investigating cellular responses during inflammation, neglecting altogether the main component undergoing physical rupture, namely, the

extracellular matrix (ECM). In a similar manner, while the regulatory role of the ECM in the progression of invasive diseases, such as cancer, is becoming acknowledged, its function in IBD is often overlooked despite the common notion that tissue destruction is imperative to disease progression.

In this article, we highlight ECM remodelling as an integral part of directional pathological signalling in IBD rather than, as is often considered, a passive bystander. We argue that the ECM, in the context of IBD, is not only a static scaffold holding cells in place and maintaining tissue architecture but a dynamic participant in intestinal immune responses capable of determining cell fate. Through our review of some common facts and new studies in the field of IBD, we will promote the new concept of the ECM immune signalling response.

WHY STUDY ECM BIOLOGY IN IBD?

Tissue damage is a hallmark of IBD progression and severity. It is known, for instance, that CD progression can advance (in some cases simultaneously) towards two seemingly opposing directions—stricturing or penetrating disease (figure 1). The stricturing type of disease results from fibrosis in the intestinal tissue,^{6–8} while the penetrating type is characterised by the formation of fistulae.⁹ Whereas the first type is a phenomenon of excess in deposition of ECM components by myofibroblasts, the second type is an outcome of ECM destruction that eliminates the boundaries between tissues. Microscopic colitis, a less common form of IBD, has an ECM component as well, the thickening of the collagenous layer in the subepithelium, which is thought to be the cause of diarrhoea in this condition.¹⁰ These examples demonstrate that ECM build-up and/or destruction play a central role in IBD, and its clinical classification and complications.

Tissue damage associated with IBD is most likely preceded by molecular events within the ECM. In figure 2, we provide evidence, for such ECM remodelling processes taking place in IBD-afflicted human colonic mucosa, revealed by second-harmonic imaging of the collagen scaffold. Second-harmonic imaging uses a two-photon microscope in order to induce an optical process in which two excitation photons are effectively combined in non-centrosymmetric materials, such as fibrillar collagen, to generate a photon with twice the excitation energy. This technique enables us to image the structure and morphology of non-labelled collagen in native tissues in high spatial resolution.

The excessive inflammatory response taking place in the intestinal tissue involves not only cellular processes but cellular processes and molecular interactions that take place in the context of a dynamically modifying ECM. Therefore, it is reasonable to assume that ECM remodelling contributes to IBD pathogenesis and inflammation sustenance and is not merely a secondary by-product.

ECM AS A PIVOTAL BIOLOGICAL ENTITY IN HOMEOSTASIS AND DISEASE

The ECM is a substantial component of tissues and thus essential for tissue function, architecture and homeostasis, yet the investigation of ECM function presents many technical challenges.

Composed of an intricate mesh of fibrous proteins and glycosaminoglycans (GAGs), the ECM serves as a scaffold for cells within tissues. It is mainly composed of three types of proteins with distinct roles: structural proteins (eg, collagen and elastin), specialised glycoproteins (eg, fibronectin) and proteoglycans (eg, lumican and decorin).¹¹ Accumulating evidence suggests that, in contrast to previous conception, the ECM is not merely a supportive platform for cells but a dynamic tissue component involved in many cellular processes, such as proliferation, migration and adhesion.¹² ECM morphology and structure are constantly undergoing remodelling; components are deposited, degraded or otherwise modified by the cues that cells convey to the matrix. In turn, this dynamic and physically variegated molecular landscape, through its interactions with cell surfaces, is able to induce directional signalling and changes in gene expression. Increased ECM stiffness has already been shown to precede fibrosis in the liver.¹³ Specifically regarding IBD with fibrostenotic phenotype, a recent study indicates that intestinal fibrosis is

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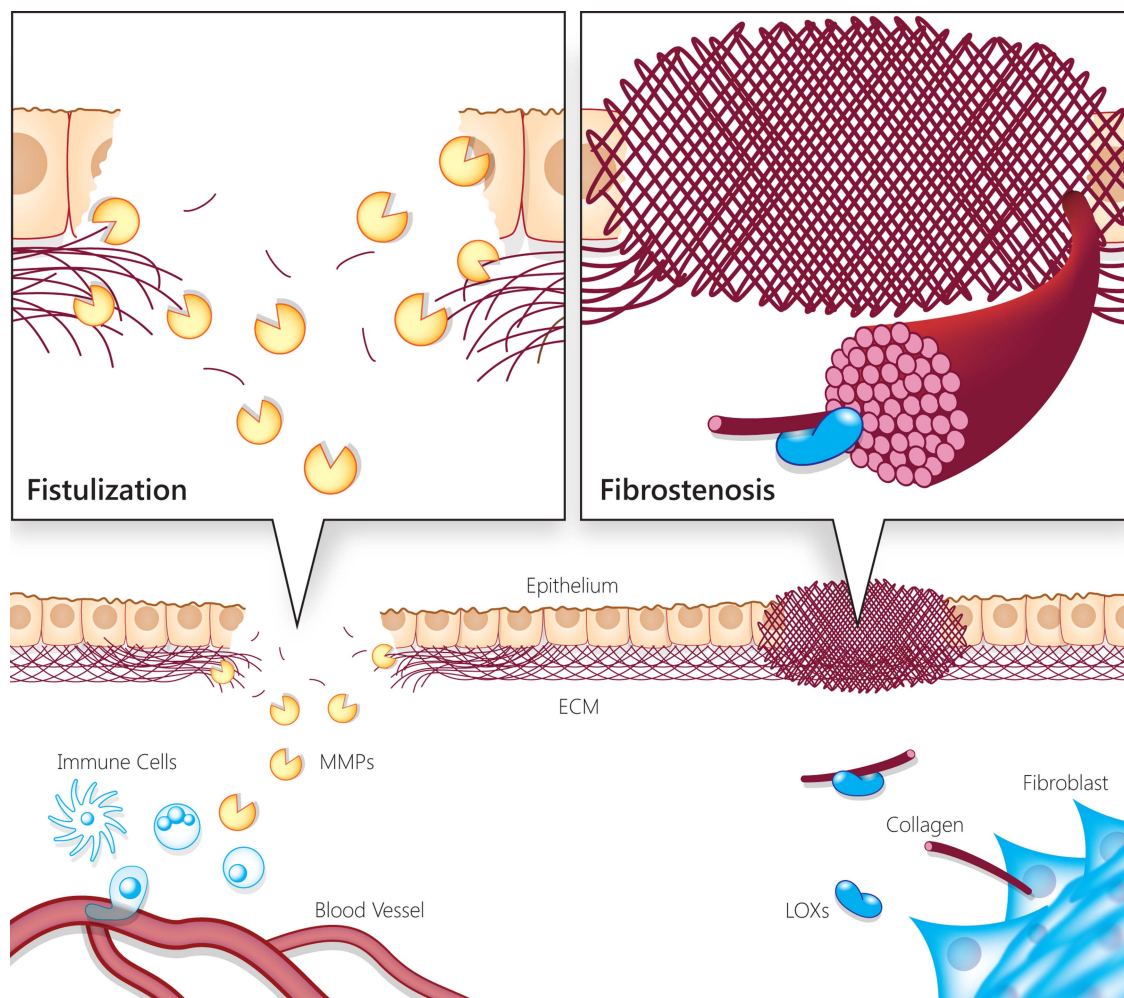


Figure 1 Illustration of IBD-associated progressive tissue damage and complications due to extracellular matrix (ECM) remodelling imbalance and dysregulation. The illustration represents the two faces of progressive tissue damage and complications associated with IBD, resulting from imbalanced and dysregulated ECM remodelling (ie, fistulising vs fibrostenotic disease). During chronic intestinal inflammation, the ECM is remodelled due to secretion of enzymes and structural components by immune, epithelial and stromal cells. Matrix metalloproteinases (MMPs) contribute to epithelial and endothelial barrier disruption and enable immune cells to infiltrate into the tissue. Extracellular proteolysis is a propagator of inflammation via cytokine processing, and release of bioactive molecules from the ECM. On the other hand, fibrotic processes also take place simultaneously by fibroblast activation and secretion of ECM components that assemble via lysyl oxidase (LOX) activity. In turn, the increased stiffness of this fibrotic tissue leads to further fibrogenesis. Therefore, both the destructive and fibrogenic processes in the ECM are self-amplifying and contribute to the tissue damage and excess inflammatory response characteristic of IBD.

autopropagated independent of inflammation, as human colonic fibroblasts cultured on stiff matrices, corresponding to CD strictures, develop a fibrogenic phenotype via mechanotransduction signalling.¹⁴

The ECM also serves as a reservoir for signalling molecules, which are exposed upon proteolysis.^{15–16} ECM remodelling processes are executed by matrix proteinases (eg, matrix metalloproteinases (MMPs) and cathepsins), lysyl oxidases (LOX/LOXL) and heparanases. These ECM remodelling enzymes are differentially expressed and contribute to various biological processes.^{17–19} In many cases, in a somewhat counterintuitive manner, enzymes with contradicting activities (eg, collagen cross-linking vs proteolysis) exhibit a synergistic effect on the same

physiological or cellular process, as demonstrated by the attenuation of breast carcinoma cell invasion following combined inhibition of LOX and MMPs.²⁰ Hence, this net enzymatic activity appears to assist the cellular crosstalk in IBD. Of note, in contrast to other post-translational modifications, the processes catalysed by ECM remodelling enzymes (eg, covalent cross-linking and proteolysis) are irreversible. This introduces a higher level of control, generating master switches in the regulation of the immune system and other physiological processes.

Accordingly, the activity of ECM remodelling enzymes is regulated on several levels, including transcription, translation, secretion, activation by cleavage of the pro-domain and inhibition by

the endogenous tissue inhibitor of metalloproteinase (TIMP) family. Thus, in the study of the ECM and its remodelling enzymes, gene expression data are far from sufficient in order to assess the actual situation in the extracellular space. This issue, along with the biochemical and physical complexity of the ECM, poses a challenge when approaching ECM involvement in biological processes including IBD pathogenesis.

ECM REMODELLING AND INTEGRITY ARE INTEGRAL FACTORS IN IBD

Various studies over the past two decades have found ECM remodelling enzymes to be upregulated in human IBD. In a complementary manner, the key role of ECM composition and remodelling in the onset,

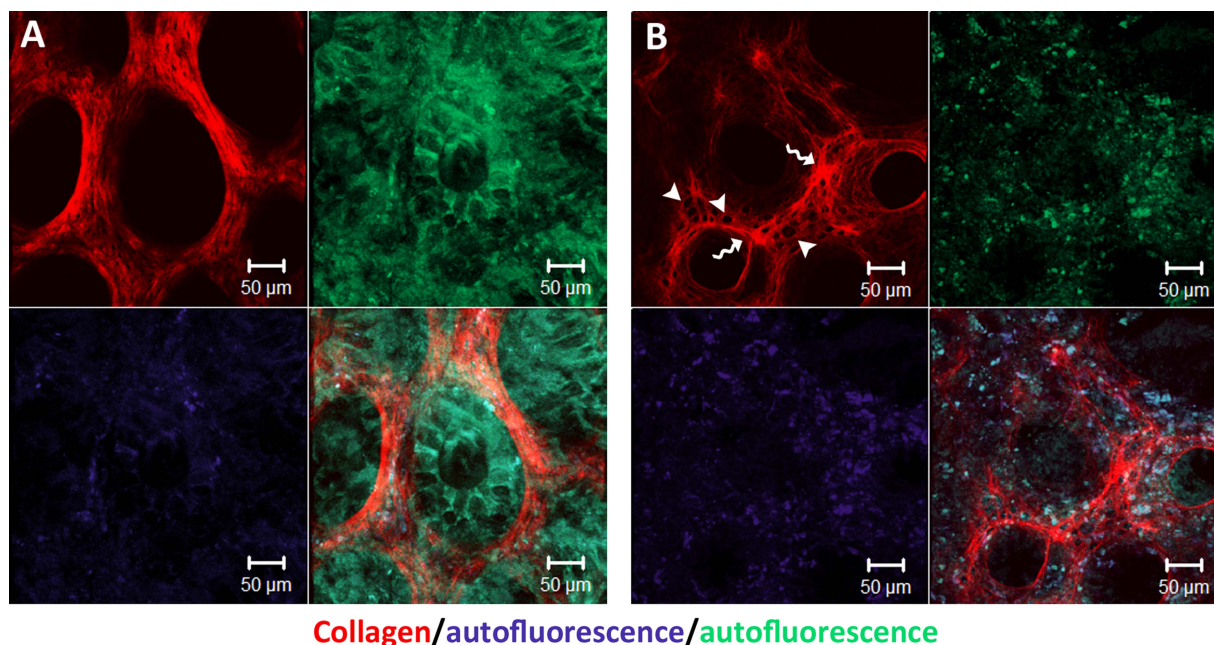


Figure 2 Second-harmonic imaging of native human colon biopsies revealing extracellular matrix (ECM) remodelling in IBD. All samples were thawed in phosphate buffered saline and immediately imaged under two-photon microscope ($\times 20$ objective). (A) Healthy colon biopsy from a patient without IBD. (B) Inflamed colon biopsy from a patient with IBD. Note the thickening of ECM barrier between crypts (indicated by curly arrows), the formation of holes within this barrier (indicated by arrowheads) and changes in collagen microstructure.

progression and severity of IBD has been implicated in various rodent models. Specifically, animal models genetically modified so as to produce an altered composition of ECM enzymes and/or components exhibit differential phenotypes in response to intestinal inflammation induction. However, the difficulty in performing mechanistic follow-up studies has left the role of ECM remodelling in IBD uncharted territory. A survey of the limited data from rodent models and human studies, presented below, reveals a clear link between ECM remodelling and IBD.

This link has been exhibited in several rodent knockout models. Heparanase, for instance, was shown to be abundant in the colonic epithelium of patients with IBD in contrast to normal colonic tissue or colonic tissue afflicted by infectious colitis,²¹ and its overexpression in mice led to an increase in colitis via enhanced macrophage activation.²² Elevated levels of collagenase-1 (MMP-1) and stromelysin-1 (MMP-3) have likewise been associated with IBD,^{23–24} as has collagenase-3 (MMP-13), which contributes to intestinal barrier dysfunction, with collagenase-3-deficient mice less susceptible to experimental colitis.²⁵ Matrilysin (MMP-7) deficiency in mice contributes to defence against luminal bacteria and re-epithelialisation,²⁶ while TIMP-3 deficiency results in higher susceptibility to experimental colitis.²⁷ However, some MMPs produce a counter-effect on disease

progression. For example, stromelysin-2 (MMP-10)-deficient mice are more susceptible to colitis and inflammation-associated colonic dysplasia.²⁸

Another revealing example of the differential function of MMPs is the highly homologous gelatinases A (MMP-2) and B (MMP-9). The former is more closely associated with homeostasis, while the latter is expressed predominantly in pathological scenarios. Interestingly, gelatinase B has been found to be the predominant upregulated MMP in inflamed intestinal tissue of patients with IBD.²⁹

Gelatinase B is secreted mainly by infiltrating neutrophils and the intestinal epithelium. Remarkably, there is a correlation between this protease's activity in the intestine and the tissue's condition, with the greatest amount exhibited in the inflamed tissue of patients with UC and CD, a lesser amount in the non-inflamed areas in the intestine of these patients, as well as intestinal tissue of patients in clinical remission, and the least amount in healthy controls. This increase in dysregulated proteolytic activity correlates with endoscopic and histological disease scores and with the extent of morphological tissue damage.^{29–33} Specifically, in fistulae of patients with CD—one of the most debilitating tissue-destructive complications of the disease—gelatinase B presents a marked increase in transcript, protein and activity levels.^{34–35}

Gelatinase B has also been recently implicated as a serological, urinary and faecal biomarker for IBD in several studies, which can be used as a tool for IBD diagnosis and monitoring. Its levels in patient bodily fluids were shown to positively correlate with other known IBD indicators and to be influenced by immunosuppressive treatments.^{36–39} Moreover, recent studies propose gelatinase B to be the superior serum biomarker for CD and UC in a group of acceptable IBD markers exhibiting a strong association with endoscopy and imaging-defined inflammation, as well as with clinical disease activity.^{24–40}

Taken together, these data suggest that gelatinase B is a reliable biomarker for human IBD, not a trivial statement considering that gelatinase B is not a canonical inflammatory molecule, such as proinflammatory cytokines. They also imply that ECM remodelling enzymes are prominent factors that characterise and strongly correlate with IBD.

The most striking findings concerning gelatinase B on disease onset demonstrate that gelatinase B-deficient mice are less susceptible to experimental colitis^{41–43} and preserve a high degree of microfloral diversity associated with protection against colitis,⁴⁴ whereas gelatinase B overexpression in the gut leads to increased susceptibility.⁴⁵ These results strengthen the view that stimulated ECM

remodelling by gelatinase B is a key mechanism in IBD pathogenesis rather than just a by-product of inflammation.

It is important to note that, in addition to their ECM maintenance duties, the structural components of the ECM also actively take part in intestinal inflammation. For example, the ECM proteoglycan Lumican was shown to contribute to innate immunity via its interaction with toll-like receptor 4 (TLR-4). Consequently, Lumican-deficient mice display a different reaction to colitis induction compared with wild-type mice, exhibiting an attenuated immune response, but also increased morbidity and tissue damage. This phenotype resembles that of myeloid differentiation primary response gene 88 (MyD88)–/– mice and TLR4–/– mice, highlighting the importance of Lumican in innate immunity.^{46–47} Thus, the enzymatic remodelling of the ECM and its composition are important in IBD development and, presumably, also its morphology and biomechanical properties.

In view of this body of evidence, it is plausible that the ECM is responsible for maintaining tissue regulation under normal conditions. Therefore, perturbation of ECM homeostatic flexibility inevitably influences the sensitivity of the intestinal tissue to inflammation. This leads us to postulate that ECM remodelling processes taking place within the intestinal tissue are indeed pivotal in IBD pathogenesis and, furthermore, are capable of steering the tissue towards health or disease.

POSTULATED IBD PATHOGENIC MECHANISMS INVOLVE ECM REMODELLING EVENTS

Several pathogenic mechanisms are thought to contribute to the chronic intestinal inflammation characteristic of IBD. Among these are (1) increased permeability of the intestinal epithelium, (2) elevated endothelial permeability and (3) self-feeding proinflammatory loops. All three mechanisms require some sort of destruction of ECM tissue barricades, which opens the way to encounters between microbial antigens and the immune system.

Changes in endothelial and epithelial barrier integrity have been associated with MMP activation, which prompts the cleavage and redistribution of cell–cell junction and adhesion proteins.^{48–54} The migration and invasion of immune cells into the gut mucosa is enabled by proteolysis of the ECM and the basement membrane, which is predominantly

performed by matrix proteases expressed by these cells.^{55–56}

In addition to their role in barrier function, certain ECM proteins have been identified as factors in inflammation. Heparanase, for example, was shown to increase macrophage sensitivity to stimulation by lipopolysaccharide, and the secretion of proinflammatory cytokines by these cells.²² Another example is the gene for LOX, which was found to be upregulated in rat colitis, and the enzyme was shown to mediate vascular inflammation induced by increased matrix stiffness.^{57–58} Gelatinase B was found to be secreted upon inflammatory stimulation, while its levels decrease in response to anti-inflammatory stimulation by interleukin (IL)-10 in a number of cell types, including immune, epithelial and stromal cells.^{59–61} Furthermore, this enzyme was shown to potentiate chemokines and cytokines such as IL-8 and IL-1 β by proteolytic processing.^{62–63}

Only one recent study suggests a specific disease-promoting mechanism for gelatinase B participation in human IBD, revealing how ECM proteolysis-generated fragments directly propagate immune signalling. This mechanism involves a proinflammatory cycle in the gut in which gelatinase B activity induces the formation of a collagen-derived fragment, proline-glycine-proline, which is a chemoattractant for neutrophils, and an inducer of gelatinase B expression.⁶⁴ This represents the first report of ECM fragment signalling in IBD pathogenesis resulting from specific proteolysis by MMPs.⁶⁵ This finding highlights the notion that elevated gelatinase B secretion to the extracellular space is not simply a result of the inflammatory process, but also an amplifier of it.

Another key mechanism indicating the regulatory role of ECM integrity in IBD pathogenesis is hyaluronan (HA)-mediated signalling. HA is an abundant GAG ECM component, and its level of polymerisation indicates matrix integrity, in contrast to fragmented HA molecules. Elevated HA fragment deposition in the intestine is associated with inflamed tissue in IBD and has been shown to induce leucocyte infiltration into the intestine and innate immune activation.^{66–67} Also, fragments of HA promote wound healing, but also fibrotic, processes by contributing to fibroblast proliferation and myofibroblast differentiation.^{68–69} Along the same lines, intact HA, formed by high-molecular-weight HA chains, promotes differentiation of anti-inflammatory regulatory T-cells.⁷⁰

In conclusion, these findings join together like pieces of a puzzle to indicate the active mechanistic role that ECM remodelling, as well as other types of extracellular proteolytic events, plays in the commonly accepted pathogenic pathways responsible for IBD development and progression.

THERAPEUTIC OPPORTUNITIES IN THE STUDY OF ECM FUNCTION IN IBD

Immunosuppressive treatments (eg, corticosteroids and antitumour necrosis factor α (TNF- α)) have had a limited effect on mucosal healing.⁷¹ Furthermore, these medications have not shown much promise in preventing or treating long-term complications of IBD (ie, fibrosis and fistulisation). There is, therefore, a pressing need to devise pharmaceutical means to prevent and treat the tissue damage in IBD.

In this respect, we now return to gelatinase B since it has been vastly explored in relation to IBD tissue damage and, as mentioned in previous sections, appears to be a prominent factor in the pathology. This notion is highlighted by several studies showing that decreased gelatinase B transcript, protein and activity levels in the intestinal tissue of patients with IBD correlate with clinical improvement and mucosal healing, facilitated by immunomodulatory treatments, such as TNF- α blockers.^{72–74} In a similar manner, anti-gelatinase neutralising antibodies developed in our laboratory exhibit great effectiveness in attenuating IBD in murine models⁷⁵ and, most recently, a humanised anti-gelatinase B antibody has entered phase I clinical trials (ClinicalTrials.gov identifier: NCT01831427). This development outlines a remarkable conceptual transformation to one that envisions the role of remodelling enzymes as therapeutic targets in IBD.

Manipulation of HA polymerisation may also be a worthy therapeutic option as this disaccharide polymer has many immunoregulatory roles. In fact, it was recently reported that the injection of high-molecular-weight HA has a beneficial effect on experimental colitis.⁷⁶

Another medical condition that may benefit from ECM remodelling factors-based therapeutics is IBD-associated fibrosis, considered an irreversible self-propagating process, currently treated mainly by mechanical means (eg, surgical resection or balloon dilation). Unravelling the molecular processes and markers preceding bowel strictures in the early inflammatory state will assist in mitigating this destructive process. As increase in matrix

stiffness seems to be an early event in tissue fibrosis, targeting collagen cross-linking enzymes, such as the LOX family, may be of therapeutic significance.

In our view, early treatment of patients with IBD consisting of a combination of immunomodulation and manipulation of ECM remodelling protocols aimed at reaching homeostatic balance in the intestinal tissue holds great promise to preventing inflammation-associated tissue damage. Furthermore, in patients already suffering from tissue destructive complications, ECM homeostatic restoration therapy may very well be the means to reverse these processes.

SUMMARY

The current outlook on IBD biology portrays ECM remodelling and processing in the pathology as a bystander effect of inflammation. We wish to challenge this notion and suggest investigating the extracellular point of view. IBD involves both tissue destruction and fibrosis. These two devastating phenomena are a direct consequence of ECM remodelling events—the first involves ECM degradation and the second, accumulating extracellular fibrogenesis. Therefore, ECM remodelling is a key event and an active participant in IBD pathophysiology.

The ECM tightly interacts with the stromal and parenchymal cells, and ensures the separation between the two groups under homeostatic conditions. Hence, any modification of the ECM influences cellular processes and, since the ECM also acts as a reservoir for signalling molecules, also induces signalling pathways. Therefore, considering that ECM biology is such a large, integral and dynamic part of the tissue, it should not be neglected when approaching any disease, especially IBD. For these reasons and in light of recent evidence from our laboratory, including the results presented herein, targeting ECM remodelling in a specific and fine-tuned manner may contribute to the treatment of IBD by preventing both propagated inflammation and tissue damage.

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REFERENCES

- Molodecky NA, Soon IS, Rabi DM, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology* 2012;142:46–54.e42; quiz e30.
- Qin X. Etiology of inflammatory bowel disease: a unified hypothesis. *World J Gastroenterol* 2012;18:1708.
- Khor B, Gardet A, Xavier RJ. Genetics and pathogenesis of inflammatory bowel disease. *Nature* 2011;474:307–17.
- Abraham C, Cho JH. Inflammatory bowel disease. *N Engl J Med* 2009;361:2066–78.
- Podolsky DK. Inflammatory bowel disease. *N Engl J Med* 2002;347:417–29.
- Rieder F, Focchi C. Intestinal fibrosis in IBD—a dynamic, multifactorial process. *Nat Rev Gastroenterol Hepatol* 2009;6:228–35.
- Latella G, Rogler G, Bamias G, et al. Results of the 4th scientific workshop of the ECCO (I): pathophysiology of intestinal fibrosis in IBD. *J Crohns Colitis* 2014;8:1147–65.
- Rieder F, de Bruyn JR, Pham BT, et al. Results of the 4th Scientific Workshop of the ECCO (Group II): markers of intestinal fibrosis in inflammatory bowel disease. *J Crohns Colitis* 2014;8:1166–78.
- Nielsen OH, Rogler G, Hahnloser D, et al. Diagnosis and management of fistulizing Crohn's disease. *Nat Clin Pract Gastroenterol Hepatol* 2009;6:92–106.
- Ingle SB, Adgaonkar BD, Hinge CR. Microscopic colitis: common cause of unexplained nonbloody diarrhea. *World J Gastrointest Pathophysiol* 2014;5:48–53. <http://www.wjgnet.com/2150-5330/pdf/v5/i1/48.pdf> (accessed 22 Jun 2014).
- Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. *J Cell Sci* 2010;123:4195–200.
- Bosman FT, Stamenkovic I. Functional structure and composition of the extracellular matrix. *J Pathol* 2003;200:423–8.
- Georges PC, Hui J-J, Gombos Z, et al. Increased stiffness of the rat liver precedes matrix deposition: implications for fibrosis. *Am J Physiol Gastrointest Liver Physiol* 2007;293:G1147–54.
- Johnson LA, Rodansky ES, Sauder KL, et al. Matrix stiffness corresponding to strictured bowel induces a fibrogenic response in human colonic fibroblasts. *Inflamm Bowel Dis* 2013;19:891–903.
- Hynes RO. The extracellular matrix: not just pretty fibrils. *Science* 2009;326:1216–19.
- Adair-Kirk TL, Senior RM. Fragments of extracellular matrix as mediators of inflammation. *Int J Biochem Cell Biol* 2008;40:1101–10.
- Page-McCaw A, Ewald AJ, Werb Z. Matrix metalloproteinases and the regulation of tissue remodelling. *Nat Rev Mol Cell Biol* 2007;8:221–33.
- Massova I, Kotra LP, Fridman R, et al. Matrix metalloproteinases: structures, evolution, and diversification. *FASEB J* 1998;12:1075–95.
- Lu P, Takai K, Weaver VM, et al. Extracellular matrix degradation and remodeling in development and disease. *Cold Spring Harb Perspect Biol* 2011;3. pii: a005058.
- Pradeep C-R, Zeisel A, Köstler WJ, et al. Modeling invasive breast cancer: growth factors propel progression of HER2-positive premalignant lesions. *Oncogene* 2012;31:3569–83.
- Waterman M, Ben-Izhak O, Eliakim R, et al. Heparanase upregulation by colonic epithelium in inflammatory bowel disease. *Mod Pathol* 2007;20:8–14.
- Lerner I, Hermanto E, Zcharia E, et al. Heparanase powers a chronic inflammatory circuit that promotes colitis-associated tumorigenesis in mice. *J Clin Invest* 2011;121:1709–21.
- Wiercinska-Drapalo A, Jaroszewicz J, Flisiak R, et al. Plasma matrix metalloproteinase-1 and tissue inhibitor of metalloproteinase-1 as biomarkers of ulcerative colitis activity. *World J Gastroenterol* 2003;9:2843–5. <http://www.ncbi.nlm.nih.gov/pubmed/14669348>
- Kofla-Dlubacz A, Matusiewicz M, Krzystek-Korpacka M, et al. Correlation of MMP-3 and MMP-9 with crohn's disease activity in children. *Dig Dis Sci* 2012;57:706–12.
- Vandenbroucke RE, Dejonckheere E, Van Hauwermeiren F, et al. Matrix metalloproteinase 13 modulates intestinal epithelial barrier integrity in inflammatory diseases by activating TNF. *EMBO Mol Med* 2013;5:932–48.
- Wilson CL, Ouellette AJ, Satchell DP, et al. Regulation of intestinal alpha-defensin activation by the metalloproteinase matrilysin in innate host defense. *Science* 1999;286:113–17.
- Monteleone I, Federici M, Sarra M, et al. Tissue inhibitor of metalloproteinase-3 regulates inflammation in human and mouse intestine. *Gastroenterology* 2012;143:1277–87.e1–4.
- Koller FL, Dozier EA, Nam KT, et al. Lack of MMP10 exacerbates experimental colitis and promotes development of inflammation-associated colonic dysplasia. *Lab Invest* 2012;92:1749–59.
- Baugh MD, Perry MJ, Hollander AP, et al. Matrix metalloproteinase levels are elevated in inflammatory bowel disease. *Gastroenterology* 1999;117:814–22.
- Bailey CJ, Hembry RM, Alexander A, et al. Distribution of the matrix metalloproteinases stromelysin, gelatinases A and B, and collagenase in Crohn's disease and normal intestine. *J Clin Pathol* 1994;47:113–16.
- Gao Q, Meijer MJW, Kubben FJGM, et al. Expression of matrix metalloproteinases-2 and -9 in intestinal tissue of patients with inflammatory bowel diseases. *Dig Liver Dis* 2005;37:584–92.
- Meijer MJW, Mieremet-Ooms MAC, van der Zon AM, et al. Increased mucosal matrix metalloproteinase-1, -2, -3 and -9 activity in patients with inflammatory bowel disease and the relation with Crohn's disease phenotype. *Dig Liver Dis* 2007;39:733–9.
- Pedersen G, Saermark T, Kirkegaard T, et al. Spontaneous and cytokine induced expression and

- activity of matrix metalloproteinases in human colonic epithelium. *Clin Exp Immunol* 2009;155:257–65.
- 34 Kirkegaard T, Hansen A, Bruun E, *et al.* Expression and localisation of matrix metalloproteinases and their natural inhibitors in fistulae of patients with Crohn's disease. *Gut* 2004;53:701–9.
- 35 Efsen E, Saermark T, Hansen A, *et al.* Ramiprilate inhibits functional matrix metalloproteinase activity in Crohn's disease fistulas. *Basic Clin Pharmacol Toxicol* 2011;109:208–16.
- 36 Manfredi MA, Zurakowski D, Rufo PA, *et al.* Increased incidence of urinary matrix metalloproteinases as predictors of disease in pediatric patients with inflammatory bowel disease. *Inflamm Bowel Dis* 2008;14:1091–6. <http://www.scopus.com/inward/record.url?eid=2-s2.0-49749130444&partnerID=40&md5=24e8f35c509c85537d05c5c0ebfbc95>
- 37 Annahazi A, Molnar T, Farkas K, *et al.* Fecal MMP-9: A new noninvasive differential diagnostic and activity marker in ulcerative colitis. *J. Crohn's Colitis*. 2012;6:316–20.
- 38 Kolho K, Sipponen T, Valtonen E. Fecal calprotectin, MMP-9, and human beta-defensin-2 levels in pediatric inflammatory bowel disease. *Int J Colorectal Dis* 2014;29:43–50.
- 39 Gao Q, Meijer MJW, Schluter UG, *et al.* Infliximab treatment influences the serological expression of matrix metalloproteinase (MMP)-2 and -9 in Crohn's disease. *Inflamm Bowel Dis* 2007;13:693–702.
- 40 Faubion WA, Fletcher JG, O'Byrne S, *et al.* EMerging BiomARKers in Inflammatory Bowel Disease (EMBARK) study identifies fecal calprotectin, serum MMP9, and serum IL-22 as a novel combination of biomarkers for Crohn's disease activity: role of cross-sectional imaging. *Am J Gastroenterol* 2013;108:1891–900.
- 41 Castaneda FE, Walia B, Vijay-Kumar M, *et al.* Targeted deletion of metalloproteinase 9 attenuates experimental colitis in mice: central role of epithelial-derived MMP. *Gastroenterology* 2005;129:1991–2008.
- 42 Garg P, Vijay-Kumar M, Wang L, *et al.* Matrix metalloproteinase-9-mediated tissue injury overrides the protective effect of matrix metalloproteinase-2 during colitis. *Am J Physiol Gastrointest Liver Physiol* 2009;296:G175–84.
- 43 Santana A, Medina C, Paz-Cabrera M-C, *et al.* Attenuation of dextran sodium sulphate induced colitis in matrix metalloproteinase-9 deficient mice. *World J Gastroenterol* 2006;12:6464–72.
- 44 Rodrigues DM, Sousa AJ, Hawley SP, *et al.* Matrix metalloproteinase 9 contributes to gut microbe homeostasis in a model of infectious colitis. *BMC Microbiol* 2012;12:105.
- 45 Liu H, Patel NR, Walter L, *et al.* Constitutive expression of MMP9 in intestinal epithelium worsens murine acute colitis and is associated with increased levels of proinflammatory cytokine Kc. *Am J Physiol Gastrointest Liver Physiol* 2013;304:G793–803.
- 46 Lohr K, Sardana H, Lee S, *et al.* Extracellular matrix protein lumican regulates inflammation in a mouse model of colitis. *Inflamm Bowel Dis* 2012;18:143–51.
- 47 Wu F, Vij N, Roberts L, *et al.* A novel role of the lumican core protein in bacterial lipopolysaccharide-induced innate immune response. *J Biol Chem* 2007;282:26409–17.
- 48 Takehara M, Nishimura T, Mima S, *et al.* Effect of claudin expression on paracellular permeability, migration and invasion of colonic cancer cells. *Biol Pharm Bull* 2009;32:825–31.
- 49 Leone AK, Chun JA, Koehler CL, *et al.* Effect of proinflammatory cytokines, tumor necrosis factor-alpha and interferon-gamma on epithelial barrier function and matrix metalloproteinase-9 in Madin Darby canine kidney cells. *Cell Physiol Biochem* 2007;19:99–112.
- 50 Xu R, Feng X, Xie X, *et al.* HIV-1 Tat protein increases the permeability of brain endothelial cells by both inhibiting occludin expression and cleaving occludin via matrix metalloproteinase-9. *Brain Res* 2012;1436:13–19.
- 51 Zuo J-H, Zhu W, Li M-Y, *et al.* Activation of EGFR promotes squamous carcinoma SCC10A cell migration and invasion via inducing EMT-like phenotype change and MMP-9-mediated degradation of E-cadherin. *J Cell Biochem* 2011;112:2508–17.
- 52 Vermeer PD, Denker J, Estin M, *et al.* MMP9 modulates tight junction integrity and cell viability in human airway epithelia. *Am J Physiol Lung Cell Mol Physiol* 2009;296:L751–62.
- 53 Qin W, Lu W, Li H, *et al.* Melatonin inhibits IL1β-induced MMP9 expression and activity in human umbilical vein endothelial cells by suppressing NF-κB activation. *J Endocrinol* 2012;214:145–53.
- 54 Behzadian MA, Wang XL, Windsor LJ, *et al.* TGF-beta increases retinal endothelial cell permeability by increasing MMP-9: possible role of glial cells in endothelial barrier function. *Invest Ophthalmol Vis Sci* 2001;42:853–9. <http://www.ncbi.nlm.nih.gov/pubmed/11222550> (accessed 16 Jun 2014).
- 55 Borregaard N, Sehested M, Nielsen BS, *et al.* Biosynthesis of granule proteins in normal human bone marrow cells. Gelatinase is a marker of terminal neutrophil differentiation. *Blood* 1995;85:812–17.
- 56 Adhikary S, Yen J-H, Ganea D, *et al.* Signaling through cannabinoid receptor 2 suppresses murine dendritic cell migration by inhibiting matrix metalloproteinase 9 expression. *Blood* 2012;120:3741–9.
- 57 Rivera E, Flores I, Rivera E, *et al.* Molecular profiling of a rat model of colitis: validation of known inflammatory genes and identification of novel disease-associated targets. *Inflamm Bowel Dis* 2006;12:950–66.
- 58 Mambetsariev I, Tian Y, Wu T, *et al.* Stiffness-activated GEF-H1 expression exacerbates LPS-induced lung inflammation. *PLoS One* 2014;9:e92670.
- 59 Gan X, Wong B, Wright SD, *et al.* Production of matrix metalloproteinase-9 in CaCO-2 cells in response to inflammatory stimuli. *J Interferon Cytokine Res* 2001;21:93–8.
- 60 Zhu YK, Liu XD, Skold CM, *et al.* Synergistic neutrophil elastase-cytokine interaction degrades collagen in three-dimensional culture. *Am J Physiol Lung Cell Mol Physiol* 2001;281:L868–78. <http://ajplung.physiology.org/content/281/4/L868> (accessed 16 Jun 2014).
- 61 Lacraz S, Nicod LP, Chicheportiche R, *et al.* IL-10 inhibits metalloproteinase and stimulates TIMP-1 production in human mononuclear phagocytes. *J Clin Invest* 1995;96:2304–10.
- 62 Van den Steen PE, Proost P, Wuyts A, *et al.* Neutrophil gelatinase B potentiates interleukin-8 tenfold by aminoterminal processing, whereas it degrades CTAP-III, PF-4, and GRO-alpha and leaves RANTES and MCP-2 intact. *Blood* 2000;96:2673–81.
- 63 Schönbeck U, Mach F, Libby P. Generation of biologically active IL-1 beta by matrix metalloproteinases: a novel caspase-1-independent pathway of IL-1 beta processing. *J Immunol* 1998;161:3340–6.
- 64 Xu X, Jackson PL, Tanner S, *et al.* A self-propagating matrix metalloprotease-9 (MMP-9) dependent cycle of chronic neutrophilic inflammation. *PLoS One* 2011;6:e15781.
- 65 Koelink PJ, Overbeek SA, Braber S, *et al.* Collagen degradation and neutrophilic infiltration: a vicious circle in inflammatory bowel disease. *Gut* 2014;63:578–87.
- 66 Kessler S, Rho H, West G, *et al.* Hyaluronan (HA) deposition precedes and promotes leukocyte recruitment in intestinal inflammation. *Clin Transl Sci* 2008;1:57–61.
- 67 Termeer C, Benedix F, Sleeman J, *et al.* Oligosaccharides of Hyaluronan activate dendritic cells via toll-like receptor 4. *J Exp Med* 2002;195:99–111.
- 68 De la Motte CA. Hyaluronan in intestinal homeostasis and inflammation: implications for fibrosis. *Am J Gastrointest Liver Physiol* 2011;301:945–9.
- 69 Webber J, Meran S, Steadman R, *et al.* Hyaluronan orchestrates transforming growth factor-beta1-dependent maintenance of myofibroblast phenotype. *J Biol Chem* 2009;284:9083–92.
- 70 Bollyky PL, Wu RP, Falk BA, *et al.* ECM components guide IL-10 producing regulatory T-cell (TR1) induction from effector memory T-cell precursors. *Proc Natl Acad Sci USA* 2011;108:7938–43.
- 71 Magro F, Rodrigues-Pinto E, Coelho R, *et al.* Is it possible to change phenotype progression in Crohn's disease in the era of immunomodulators? Predictive factors of phenotype progression. *Am J Gastroenterol* 2014;109:1026–36.
- 72 Makitalo L, Sipponen T, Karkkainen P, *et al.* Changes in matrix metalloproteinase (MMP) and tissue inhibitors of metalloproteinases (TIMP) expression profile in Crohn's disease after immunosuppressive treatment correlate with histological score and calprotectin values. *Int J Colorectal Dis* 2009;24:1157–67.
- 73 Geboes K, Rutgeerts P, Opdenakker G, *et al.* Endoscopic and histologic evidence of persistent mucosal healing and correlation with clinical improvement following sustained infliximab treatment for Crohn's disease. *Curr Med Res Opin* 2005;21:1741–54.
- 74 De Bruyn M, Arijis I, Wollants W-J, *et al.* Neutrophil gelatinase B-associated lipocalin and matrix metalloproteinase-9 complex as a surrogate serum marker of mucosal healing in ulcerative colitis. *Inflamm Bowel Dis* 2014;20:1198–207.
- 75 Sela-Passwell N, Kikieri R, Dym O, *et al.* Antibodies targeting the catalytic zinc complex of activated matrix metalloproteinases show therapeutic potential. *Nat Med* 2012;18:143–7.
- 76 Chen H, Mahaseth M, Zhang Y. Hyaluronic acid as a rescue therapy for trinitrobenzene sulfonic acid-induced colitis through Cox-2 and PGE2 in a Toll-like receptor 4-dependent way. *J Zhejiang Univ Sci B* 2011;12:712–19.