		spontaneo	us resolvers	acute	elv infected	chro	nic patients
number of patie	nts	16	%	16	%	108	%
sex	female	5	31.3	7	43.8	31	28.7
	male	11	68.8	9	56.3	77	71.3
	•						
Age	>40	7	43.8	5	31.3	35	32.4
	<40	9	56.3	11	68.8	73	67.6
Ethnicity	Caucasian	16	100.0	15	93.8	87	80.6
	Asian					16	14.8
	Hispanic			1	6.3	1	0.9
	not known					4	3.7
Risk factor	IVDU	11	68.8	9	56.3	70	64.8
	unkown	1	6.3			14	13.0
	BP			1	6.3	12	11.1
	cocaine	-				6	5.6
	tattoo	2	12.5	1	6.3	3	2.8
	needlestick	1	6.3	_		2	1.9
	sexual	1	6.3	5	31.3	0	0.0
	vertical					1	0.9
	Construine 2		NI/A	0	50.0	14	12.0
nev subtype	Genotype 3		N/A	8	50.0	14	13.0
	Genotype 3a		IN/A	0	50.0	94	07.0
Viral load	III/ml (mean)		N/A		1827490		872828
Viral load	III/ml (range)		N/A		6290-8930000		3614-17880410
	io, iii (idiigo)		.,,,,		0230 0350000		5011 1/000110
Outcome	cleared spontaneously	16	100.0	4	25.0	0	0.0
	chronic	0	0.0	10	62.5	108	100.0
	not treated	N/A		6	60.0	30	27.8
	treated	N/A		4	40.0	78	72.2
	unkown			2	12.5		
	ч		•				
Treatment	SVR			2	50.0	42	53.8
	REL			1	25.0	25	32.1
	NR			1	25.0	6	7.7
	incomplete					3	3.8
	not known					2	2.6

# Table S1: Patient details of patients with spontaneously resolved, acute and chronic HCV genotype-3 infection.

ID: patient identifier; IVDU: intravenous drug use; BP: blood products; SVR: sustained virological response; REL: viral relapse; NR: treatment non-response.

ID	Age	Sex	Race	Risk factor	GT	HCV VL	ALT	Treatment	outcome
						lu/ml			
A1	23	F	W	Sexual	3	>700000	68	naïve	cleared spontaneously
A2	31	М	W	IVDU	3a	6290	not known	naïve	cleared spontaneously
A3	23	M	W	IVDU	3a	8930000	46	naïve	cleared spontaneously
A4	21	М	W	IVDU	3	15900	72	naïve	cleared spontaneously
A5	19	М	W	IVDU	3	168000	229	naïve	chronic
A6	20	F	W	IVDU	3	16100	38	naïve	chronic
A7	46	M	W	blood exposure	3	204000	592	naïve	chronic
A8	28	M	W	Sexual	3	136000	698	naïve	chronic
A9	45	F	W	Tattoo	3a	>700000	201	naïve	chronic
A10	21	F	W	IVDU	3a	600000	not known	naïve	chronic
A11	18	F	W	IVDU	3	102000	88	naïve	lost in <6 months
A12	25	F	W	IVDU	3a	not known	21	naïve	lost in <6 months
A13	40	F	W	Sexual	3a	>700000	464	treated	cleared on Tx
A14	41	M	Н	Sexual	3a	67535	250	treated	cleared on Tx
A15	49	М	W	Sexual	3a	46603	1441	treated	cleared on Tx
A16	20	М	W	IVDU/Tattoo	3	>700000	383	treated	chronic (non-responder)

#### Table S2: Patient details acute HCV genotype-3 cohort.

ID: patient identifier; M: male; F: female; W: white; H: Hispanic; IVDU: intravenous drug use; GT: genotype; ALT: alanine aminotransferase; Tx: treatment.

Protein	HLA	Wildtype	Variant	Amino Acids	Peptide ID	Predicted Epitope
NS2	A0101	V	А	879-887	001-A0101	VILLTSLLY
	A02	L	Р	862-871	049-A02	ALQVWVPPL <b>L</b>
		L	Р	870-879	048-A02	L <b>L</b> ARGSRDGV
		Y	Н	881-890	052-A02	LLTSLLYPSL
		Y	Н	885-894	047-A02	LLYPSLIFDI
	A0301	V	А	926-935	006-A0301	RLCMLVRS <b>V</b> M
		V	А	929-938	004-A0301	MLVRS <b>V</b> MGGK
		V	А	930-939	005-A0301	LVRS <b>V</b> MGGKY
	A2402	Y	Н	886-894	007-A2402	LYPSLIFDI
	B1501	Т	Α	878-887	008-B1501	GVILLTSLLY
	B15	Ι	V	946-954	055-B15	SIGRWFNTY
	B2705	R	K	940-949	010-B2705	FQMIILSIG <b>R</b>
		R	K	948-956	009-B2705	G <b>R</b> WFNTYLY
		Н	Y	962-971	011-B2705	MQ <b>H</b> WAAAGLK
	B4402	Ι	V	822-831	012-B4402	ATLGAG <b>I</b> LVL
	B44	Ι	V	826-835	057-B44	AGILVLFGFF
	B5101	S	G	871-880	013-B5101	LARG <b>S</b> RDGVI
	C03	L	М	829-838	050-C03	I <b>L</b> VLFGFFTL
	C04	V	Ι	981-990	051-C04	IFSPMEIK <b>V</b> I
NS3	A0101	Y	F	1442-1450	014-A0101	ATDALMTGY
	A0201	А	D	1389-1398	015-A0201	ALLKGGRHLI
	A0301	V	Ι	1138-1146	016-A0301	L <b>V</b> TRDADVI
	B1501	K	R	1296-1305	017-B1501	<b>K</b> LTYSTYGKF
	B2705	L	Ι	1379-1388	018-B2705	IPFYGKAIP <b>L</b>
		V	Ι	1632-1641	019-B2705	YRLGP <b>V</b> QNEI
	B4402	G	S	1407-1416	020-B4402	DEIASKLR <b>G</b> M
	B4403	L	S	1639-1647	021-B4403	NEICLTHPI
	B5101	А	D	1388-1397	023-B5101	IALLKGGRHL
NS4B	A02	А	Т	1873-1882	062-A02	KIMGGELPT <b>A</b>
		А	Т	1880-1889	065-A02	PTAEDMVNLL
	A0301	Ι	V	1901-1910	026-A0301	GVICAAILRR
	A2601	R	K	1852-1861	027-A2601	RVLLDILAGY
	A68	А	Ι	1736-1744	067-A68	EK <b>A</b> LGLLQR
	B27	R	K	1948-1957	061-B27	ARVTALLSSL
	B4001	Ι	V	1847-1855	028-B4001	GIGLGRVLL
	24	А	Т	1879-1888	066-B51	LPT <b>A</b> EDMVNL
	B51	А	Т	1881-1889	064-B51	TAEDMVNLL
	C0401	V	Ι	1733-1742	025-C0401	QFKEK <b>V</b> LGLL
NS5A	A0201	G	S	2321-2330	029-A0201	ALPPR <b>G</b> APPV
	A0301,	V, P	A, S	2382-2391	030-A0301	K <b>V</b> PPS <b>P</b> GGES
	A2601	D	G	2268-2276	033-A2601	ETDAELSVA
	A68	Т	S	1989-1998	070-A68	WVC <b>T</b> VLSDFK
	B0702	V	Е	2309-2317	036-B0702	APDY <b>V</b> PPTV
		V	Е	2313-2322	035-B0702	<b>V</b> PPTVHGCAL
		Т	А	2332-2341	034-B0702	PPRRKR <b>T</b> IQL
	B0801	Ι	V	2251-2259	037-B0801	ESETKVVIL
	B44	R	Q	2097-2105	071-B44	VEVR <b>R</b> VGDF
NS5B	A0201	Т	I	2489-2498	038-A0201	VLDDHYK <b>T</b> AL
		Ν	D	2540-2549	040-A0201	SLSSKAI <b>n</b> qi
		Ν	D	2544-2552	039-A0201	KAI <b>N</b> QIRSV
	A1101	R	K	2500-2509	041-A1101	EVKERAS <b>R</b> VK
	A2601	K	R	2537-2545	042-A2601	DVRSLSS <b>k</b> A
	B0801	Ι	М	2507-2515	043-A2601	RVKARMLTI
	B1501	Q	L	2476-2484	044-B1501	S <b>q</b> rqkkvtf
	B5101	ĸ	R	2474-2482	045-B5101	SASQRQ <b>K</b> KV

# Table S3: HLA-associated polymorphisms – sequence polymorphisms located

*within* the predicted epitope sites. HCV viral region, associated HLA type, epitope position, peptide identifier and sequence, as well as wild-type and variant sequences are depicted. Polymorphic sites associated with patient HLA are marked in in bold.

Protein	HLA	Wildtype	Variant Residue	Amino Acids	Polymor- phic Site	Peptide ID	Predicted Epitope and Wild-type Residue ()
NS2	A0201	Ι	V	934-942	943	002-A0201	VMGGKYFQM( <b>I</b> )
		Ι	V	944-953	943	003-A0201	(I) ILSIGRWFNT
	A02	L	Р	861-870	871	054-A02	AALQVWVPPL( <b>L</b> )
		Y	Н	878-886	887	053-A02	GVILLTSLL (Y)
NS3	A68	L	F	1047-1056	1046	058-A68	(L)GTIVTSLTGR
	B5101	Α	D	1379-1388	1389	022-B5101	IPFYGKAIPI ( <b>A</b> )
		Α	D	1390-1398	1389	024-B5101	(A) QLKGGRHLI
NS5A	A1101	V	Α	2374-2382	2383	031-A1101	DTQSSTTSK ( <b>V</b> )
	A2402	D	Е	2138-2147	2148	032-A2402	RYAPPCKPLL ( <b>D</b> )
NS5B	B5101	М	L	2846-2854	2855	046-B5101	APTIWVRMV( <b>M</b> )

## Table S4: HLA-associated polymorphisms – sequence polymorphisms located *flanking* the predicted epitope sites.

HCV viral region, associated HLA type, epitope position, peptide identifier and sequence as well as wild-type and variant sequences are depicted. Polymorphic sites flanking epitope are indicated in bold within brackets ().

Protein	Position	3a peptide sequence	Patient	Patient viral sequence	CD8/CD4
core	27-51	GGQIVGGVYVLPRRGPRL	C58 *	GGQIVGGVYVLPRRGPRL	N/A
		VYVLPRRGPRLGVRATRK	C106	VYVLPRRGPRLGVRATRK	
			S11	SR	
	66-90	PKARRSE GRSWAQPGYPW	C5	ND	CD4
		GRSWAQPGYPWPLYGNEG	C12	PKARRSEGRSWAQPGYPW	
			C37	PKARRSEGRSWAQPGYPW	
			450	PKARRSEGRSWAQPGYPW	
			C68 *	GRSWAQPGYPWPLYGNEG	
1 1	130-147	FADLMGYIPLVGAPVGGV	C15	FADLMGYIPLVGAPVGGV	N/A
			C5	ND	
	137-154	IPLVGAPVGGVARALAH	S15	SR	N/A
	143-158	PVGGVARALAHGVRAL	A16	PVGGVARALAHGVRAL	CD4
			C6 *	PAGGVARALAHGVRAL	
			C12	PVGGVARALAHGVRAL	
			C13	PVGGVARALAHGVRAL	
			C18	PVGGVARALAHGVRAL	
			C19	PVGGVARALAHGVRAL	
			C22	PVGGVARALAHGVRAL	
			C23	PVGGVARALAHGVRAL	
			C27	PVGGVARALAHGVRAL	
			331 *	PVGGVARALAHGVRAL	
			C70	PVGGVARALAHGVRAL	
			C77 *	PVGGVARALAHGVRAL	
ľ	148-165	ARALAHGVRALEDGINFA	C103	ARALAHGVRALEDGINFA	CD8
E2	460-476	CKPITEFROGWGSLTDA	A15	CKPITFFROGWGSLTDA	CD4
		FROGWGSLTDANTTGPSD	C106	FNOGWGSLTDANT SGPSD	
	610-625	LTPRCMVDYPYRLWHY	C106	LTPRCLVDYPYRLWHY	N/A
			S1 *	SR	,
			S12	SR	
1 1	635-650	KVRMFVGGFEHRFTAA	A11	KVRMFVGGFEHRFTAA	N/A
			C13	KVRMFVAGFEHRFTAA	,
1 1	696-719	LIHLHONIVDVQYLYGV	S4 *	SR	CD8
		NIVDVQYLYGVGSGMVGW	A16	NIVDVQYLYGVGSGMVGW	
			S9 *	SR	

#### Table S5: HCV genotype-3-specific T-cell targets in HCV structural regions.

HCV genotype-3-specific T-cell targets identified using overlapping peptide pools in HCV structural regions are depicted. All patients targeting specific individual peptides are specified (colour coding: orange-SR, blue-acute, yellow chronic). Circulating viral sequence is depicted when obtained, as well as CD4+ /CD8+ subset analysis. Sequence polymorphisms differing from overlapping peptide set sequence are marked in red. \* Epitopes previously described in (Humphreys et al. 2012). N/A not available.

Protein	Position	3a peptide sequence		Patient		Patient viral sequence	CD8/CD4
NS2	886-896	(L) LYPSLIFDI	#	A8		LLYPSLIFDI	CD8
			#	A8		LYPSLIFDI	
			#	C1		LYPSLIFDI	
			#	C19		LYPSLIFDI	
			#	C47		LYPSLIFDI	
	001 010		#	C58		LY <mark>S</mark> SLIFDI	
	931-940	LVRSVMGGKY	#	\$15		SR	CD8
	941-951	FQMIILSIGR	#	A16		FQMAILSIGR	CD8
			#	C38		FQMIIL <b>GV</b> GR	
			#	C/3		FQMVILSIGK	
NS3	1040-1062	AQQTRGLLGTIVTSLTGR		S15		SR	N/A
		LGTIVTSLTGRDKNVV					
	1139-1147	LVTRDADVI	#	S15		SB	CD8
	1198-1213	KALOFT BUETL STOAR		\$5	*	SB	N/A
	1246-1261	KUPAAVVAOGYNULUI.		58	*	SR	N/A
	1264-1281	SVAATLGEGSEMSRAVGT		A11		SVAATLGEGSEMSHAYGI	N/A
	1201 1201	ovinitizer corribititier		56		SB	,
	1282-1305			S1	*	SR SR	N/A
	1202-1303	CND WINTER CARL WARMAN		512		SR	N/A
	1270 1207	GNRTVITGARLTISTIGR		A12		SR	CD9
	1370-1387	LEVALGSEGEIPFIGRAI	4	A13		ND	CDo
	1379-1307	IFFIGRATEI	π 4	A13		ND	
	1423-1440		Ť	C22			N/A
	1423-1440	ATTRGLDVSVIPIAGDVV		C22	*	ATTRGLDVSVIPIAGDVV	
	1436-1447	CDURINGARDALMECE		A15		SK CDUURCARDAI MRCY	CD4
	1430-1447	GDVVVCAIDALMIGF	4	A15		AMDALMICI	CD0
	1442-1447	AIDALMIGI	π #	C68			
			π #	C106		ATDALMIGE	
			" #	C108		ATDALMTCE	
	1520-1537	RESOMEDSVVLCECYDA	π	C5		ND	CD8
	1020 1007	DSVVLCECYDAGCSWYDL		C7		RPSGMEDSVVLCECYDAGCSWYDL	020
				C15	*	RPSGMEDSVVLCECYDAGCSWYDL	
				C17		RPSGMFDSVVLCECYDAGCSWYDL	
				C19		RPSGMFDSVVLCECYDAGCSWYDL	
				C27	*	RPSGMFDSVVLCECYDAGC AWYDL	
				C35		ND	
				C42		ND	
				C44		RPSGMFDSVVLCECYDAGCSWYDL	
				C58	*	RPSGMFDSVVLCECYDAGCSWYDL	
				C96		ND	
				C98		ND	
				S13		SR	
				S14		SR	
	1547-1569	RAYLSTPGLPVCQDHLDF		C19		RAYLSTPGLPVCQDHLDFWESVF	N/A
		GLPVCQDHLDFWESVF					
NS4B	1792-1808	PAVASLMAFTASVTSPL		A8		ND	CD8
				C68	*	PAVASLMAFTASVTSPL	
				C77	*	ND	
	1805-1822	TSPLTTNQTMFFNILGGW		S8	*	SR	N/A
				S13		SR	
	1825-1842	THLAGPQSSSAFVVSGLA		C77	*	ND	N/A
	1853-1862	RVLLDILAGY	#	A16		<b>K</b> VLLDILAGY	CD8
			#	A4		ND	
			#	C5		<b>K</b> VLLDILAGY	
			#	C52		ND	
			#	C77		ND	
	1917-1932	EGAVQWMNRLIAFASR		C67	*	EGAVQWMNRLIAFASR	CD8
NS5A	2030-2047	GVMSTRCPCGASTAGHVK		C13	*	GVMSTRCPCGAST TGHVK	CD8
	2119-2136	CPCOVPAAEFFTEVDGVR		A8		CPCOVPA PEFFTEVDGVR	000
	2126-2141	AEFETEVDGVRLHRYA		A13		ND	CD8
				A15		ND	
				S10		SR	
				S15		SP	
	2145-2162	KPLURDETTEMVGLNSYA		A5		ND	N/A
	2404 0 100				_	-142	
NS5B	2484-2499	TFDRLQVLDDHYKTAL		A13		TFDRLQVLDDHYKTAL	CD8
	2490-2499	VLDDHYKTAL	#	C68		ND	
	2508-2516	RVKARMLTI	#	A13		RVKARMLTI	CD8
			#	S10		SR	
	2548-2565	NQIRSVWEDLLEDTTTPI		S2	*	SR	CD4
	2603-2618	KRALYDVIQKLSIETM		S11	*	SR	CD4
	2844-2861	IMYAPTIWVRMVMMTHFF		A4		ND	N/A
	2893-2908	IIERLHGLSAFTLHSY		S2	*	SR	CD4
	2947-2964	GKAKICGLYLFNWAVRTK		C37	*	GKAKI <b>T</b> GLYLFNWAVRTK	CD8
1	2967-2976	KLTPLPAAGQL		450	*	KLTPLPAAG <b>L</b> L	CD8

**Table S6: HCV genotype-3-specific T-cell targets in HCV non-structural regions.** HCV genotype-3-specific T-cell targets identified using overlapping peptide pools and HLA predicted peptides (#) in HCV structural regions are depicted. All patients targeting specific individual peptides are specified (colour coding: orange-SR, blueacute, yellow chronic). Circulating viral sequence is depicted when obtained, as well as CD4+ /CD8+ subset analysis. Sequence polymorphisms differing from overlapping peptide set sequence are marked in red. \*Epitopes previously described in (Humphreys et al. 2012). N/A not available.

		G	GT3 CD8+ epitopes (Oxford Study Cohort)						GT1 CD8+ epitopes in corresponding regions			
	Position	3a peptide sequence	Viral protein	HLA	Pt ID	HLA type				Peptide (Literature)	HLA	First author
	931-940	LVRSVMGGKY	NS3	A03	S15	A*3201 A*0301 B*14	01 B*0702 C*080	2 C*0702	#	no CD8 epitopes describe	d	
	941-951	FQMIILSIGR	NS2	B27	A16 C38 C73	A*0201 A*2601 B*22 A*0201 A*1101 B*18 A*0201 B*52	02 B*3801 C*120 01 B*2705 C*010 01 B*2702 C*070	3 2 C*1203 1 C*1501	# # #	no CD8 epitopes describe	d	
	1917-1932	EGAVQWMNRLIAFASR	NS4B		C67	A*0101 A*3001 B*13	02 B*4402 C*060	2 C*0501	# *	no CD8 epitopes describe	d	
	2030-2047	GVMSTRCPCGASIAGHVK	NS5A		C13	A*1101 A*7401 B*44	03 B*38 C*04	C*0702	# *	no CD8 epitopes describe	d	
	2484-2499	TEDRLOVI.DDHYKTAI.	NS5B		A13	A*0101 B*08	01 B*5101 C*010	2 C*0701	#	no CD8 epitopes describe	d	
	2490-2499	VLDDHYKTAL		A02	C68	A*0101 A*0201 B*08	01 B*5701 C*060	2 C*0701	#			
	2508-2516	RVKARMLTI	NS5B	B08	A13	A*0101 B*08	01 B*5101 C*010	2 C*0701	#	no CD8 epitopes describe	d	
					\$10	A*3201 A*0101 B*08	01 B*4402 C*050	1 C*0701				
	2967-2976	KLTPLPAAGQL	NS5B		450				# *	no CD8 epitopes describe	d	
-	1853-1862	RVLLDILÄGY	NS4B	A26	A16 A4 C6 C52 C77	A*0201 A*2601 B*27 A*0201 A*2601 B*38 A*2301 A*2601 B*38 A*0101 A*2601 B*38 A*0101 A*2601 B*38	02         B*3801         C*120           01         B*4402         C*050           01         B*4901         C*070           01         B*0801         C*060           01         B*2702         C*120	301 1 C*1203 2 C*0701 3 C*0102	# # # #	ILAGYGAGV ILAGYGAGV ILAGYGAGV	A2 A2 A2	M Battegay N H Gruener T Kuntzen
	886-896	LLYPSLIFDI/LYPSLIFDI	NS2	A02/A24	A8 C1 C19 C47 C1 C58	A*2402         A*0201         B*31           A*1101         A*2404         B*11           A*24         B*33           A*0205         A*2402         B*11           A*0205         A*2402         B*11           A*0205         A*2402         B*11           A*0101         A*0201         B*01	602         B*4403         C*040           B*3501         C*040           601         B*4403         C*040           802         B*4901         C*060           802         B*4901         C*060           802         B*4901         C*060           802         B*4001         C*030	1 C*1601 1 C*1203 1 C*0409 2 C*0701 2 C*0701 4 C*0702	# # # #	<u>HPTLVFDI</u> TK HPTLVFDITKL	Class I Class I	A L Cox T Kuntzen
	2947-2964	GKAKICGLYLFNWAVRTK	NS5B		C37	A*1101 B*0	'02 B*4402 C*050	1 C*0702	# *	RGGRAAICGKYLFNWAVR GRAAICGKY GRAAICGKYLFNWAV KYLFNWAVK	Class I B27 Class I A2	C Neumann-Haefelin C Neumann-Haefelin P T F Kennedy Z Guo
ł	148-165	ARALAHGVRALEDGINFA	core		C103				#	GVRVLEDGV	A2	H F Löhr
										RVLEDGVNY VLEDGVNYATGNLPG	Class I Class I	D D Anthony K Sugimoto
-	1139-1147	LVTRDADVI	NS3	A03	975	A*3201 A*0301 B*14	01 B*0702 C*080	2 C*0702	#	VTRHADVIPV	Class I	T Kuntzen
ļ	1520-1527	DROWEDOURU OROVDA COOLUDI	NC2	P25/C04	CE	A*1101 A*1101 P*20	01 8*5101 C*040	1 C*1402		MEDOQUILOBOVD300	Class I	D. Ciuffrodo
	1520-1537	RESCREDS VULLEL TURCS WITH		635/C04	C3 C7 C15 C17 C19 C27 C35 C42 C44 C58 C96 C98 S13 S14	A 1101 A 1101 B 1 A 1010 A 10201 B 73 A 72402 A 3002 B 70 A 1101 A 1101 B 11 A 1101 A 1101 B 11 A 1101 A 2402 B 14 A 1011 A 2402 B 73 A 2402 B 73 A 2402 B 73 A 2402 B 73 A 2402 A 1101 B 74 A 2402	101         B 5101         C*040           102         B 3501         C*040           117         B 3501         C*040           118         B 3501         C*040           110         B 3501         C*040           111         B 5501         C*040           111         B 4403         C*040	1 C*1402 1 C*0802 1 C*0701 1 C*1203 1 C*0702 1 C*1601 1 C*0409 1 C*0409 1 C*0301 1 C*0202 1 C*0501	# * # * # *	REUSSVILLUTURE.		
	2126-2141	AEFFTEVDGVRLHRYA	NS5A		A13	A*0101 B*08 A*0101 B*55	01 B*5101 C*010	2 C*0701	#	FFTELDGVRLHRFAP	Class I	D Ciuffreda
					S10	A*3201 A*0101 B*08	01 B*4402 C*050	1 C*0701				
			101-		315	A 3201 A-0301 B+14	01 D-0702 C*080	2 0.0702		-		
	1/92-1808	PAVASLMAFTASVTSPL	NS4B		6-23 C68	A*2402 A*0201 B*35 A*0101 A*0201 B*08	02 B*4403 C*040 01 B*5701 C*060	1 C*1601 2 C*0701	# *	SLMAFTAAV SLMAFTAAV	A2 A2	B Rehermann K M Chang
					C//	A-UZUI A*2601 B*38	DUI B*2/02 C*120	s C*0102	# *	SLMAFTAAV	AZ	N H Gruener
	696-719	LIHLHQNIVDVQYLYGVGSGMVGW	E2		S4 A16 S9	A*02 A*3201 B*23 A*0201 A*2601 B*23	07 B*1501 C*060 02 B*3801 C*120	2 C*0304 3	# * # *	ALSTGLIHLHQNIVD LHQNIVDVQYLYGVG	Class I Class I	D Ciuffreda D Ciuffreda
	1379-1387	IPFYGKAIPI	NS3	B51	A13	A*0101 B*08	01 B*5101 C*010	2 C*0701	#	IPFYGKAI &	B51	S Giugliano
					AZ	B*44	HUS B*5101 C*140	2 C*1601	Ŧ	1PFYGKAIPL	851	U teny
	1436-1447 1442-1447	GDVVVCATDALMTGF ATDALMTGY	NS3	A01	A15 A15 C108 C106 C68	A*0101         B*55           A*0101         B*55           A*0101         A*2402           B*0101         A*2402           A*0101         A*2402           B*0101         A*0301           B*0101         A*0201           B*0101         A*0201	201 B*5701 C*060 201 B*5701 C*060 301 B*3906 C*070 702 B*0702 C*070 301 B*5701 C*060	2 C*1202 2 C*1202 1 C*0702 2 C*0702 2 C*0702 2 C*0701	# # #	ATDALMTGY VATDALMTGY ATDALMTGF & & ATDALMTGF ATDALMTGF ATDALMTGY ATDALMTGY ATDALMTGY	A1 Class I A1 A1 A1 A1 A1 A1	G M Lauer, 2002 A M Werthheimer S Giugliano T Kuntzen A L Cox E Barnes G M Lauer, 2004

### Table S7: Overlap between genotype-3-specific CD8+ T-cell targets defined in this study and previously described HCV genotype-1-specific CD8+ targets.

HCV genotype-3-specific CD8+ epitopes and described genotype-1-specific CD8+ epitopes were classified by overlap (left coloured bar): 'overlapping' (blue), 'likely overlapping' (light blue, <20% sequence differences within targeted area), 'unlikely overlapping' (light red, >20% differences within targeted area, or less than 7 amino acids overlap), and 'not overlapping' (red). *Left:* For each epitope, viral region, position and genotype-3 sequence is given, as well as patients targeting the epitopes (colour coding: orange-SR, blue-acute, yellow chronic), and patient's HLA class-I types. *Right:* Published overlapping genotype-1 CD8+ epitopes are specified, including sequence, restricting HLA type and first author of the publication; overlap between epitopes is underlined and genotype-3/genotype-1 sequence differences marked bold. # CD8 restriction for marked patient experimentally defined; \* Epitopes

previously described in (Humphreys et al. 2012); & Epitope previously described for genotype-1 and 3.

(M Battegay et al. 1995; Rehermann et al. 1996; K. M. Chang et al. 1997; Löhr et al. 1999; Grüner et al. 2000; Wertheimer et al. 2003; Sugimoto et al. 2003; Lauer et al. 2004; Cox et al. 2005; Kennedy et al. 2006; Kuntzen et al. 2007; Yerly et al. 2008; Ciuffreda et al. 2008; Giugliano et al. 2009; Neumann-Haefelin et al. 2008; Barnes et al. 2012; Guo et al. 2012)

	GT3 CD4+ epit	opes (O		GT1 epitopes described in corresponding regions					
		Viral	Patient						
Position	3a peptide sequence	protein	ID				Peptide (Literature)	HLA	First author
453-476	CPQRLSSCKPITFFRQGWGSLTDANITGPSD	E2	A15	DRB1*0701	DRB1*1502	#	no CD4 epitopes described		
			C106						
				_					
2603-2618	KRALYDVIQKLSIETM	NS5B	S11	DRB1*0404	DRB1*0701	# *	KL <b>PLAV</b> MGSSYGFQYSPGQR	Class-II	J Schulze zur Wiesch, 2007
							KL <b>PLAV</b> MGSSYGFQYSPGQR	Class-II	C L Day
							1		
66-90	PKARRSEGRSWAQPGYPWPLYGNEG	core	C5	DRB1*0101	DRB1*0701		RRQPIPKARRPEGRTWAQPG	Class-II	J Schulze zur Wiesch, 2012
			C12	DRB1*0301	DRB1*0401		RQPI <u>PKVRR<b>P</b>EGR<b>T</b></u>	HLA-DR	] ] Lasarte
			C37	DRB1*0401	DRB1*0407		KVRRPEGR TWAQPG	HLA-DR	] ] Lasarte
			450			#	PEGRTWAQPGYPWPLYGNEG	Class-II	J Schulze zur Wiesch, 2007
			C68	DRB1*0101	DBR1*0701	# *	PEGRTWAQPGYPWPLYGNEGCGW	Class-II	H F Löhr
							PEGRTWAQPGYPWP	HLA-DR	J J Lasarte
				_			PEGRTWAQPGYPWPL	Class-II	P T F Kennedy
143-158	PVGGVARALAHGVRAL	core	A16				ADLMGYIPLVGA PLGGAARA	Class-II	J Schulze zur Wiesch, 2007
			C6	DRB1*0801	DRB1*1101	# *	ADLMGYIPLVGAPLGGAAR	HLA-DR	F A Castelli
			C12	DRB1*0301	DRB1*0401	#	ADLMGYIPLVGAPLGGAARA	Class-II	J Schulze zur Wiesch, 2012
			C13	DRB1*1504	DRB1*0701		LMGYIPLVGAPLGGA	Class-II	D Ciuffreda
			C18	DRB1*0301	DRB1*1602	#	LVGAPLGG <b>A</b> ARAL	Class-II	H F Löhr
			C19	DRB1*0403	DRB1*0701		LVGAPLGG <b>A</b> ARALAH		K Sugimoto
			C22	DRB1*0404	DRB1*1104	#	GAP <b>L</b> GG <b>A</b> ARALAHGVR <b>V</b> LED	Class-II	J Schulze zur Wiesch, 2007
			C23	DRB1*0701	DRB1*0801	#	GAPLGGAARALAHGVRVLED	Class-II	A J MacDonald
			C27	DRB1*0101	DRB1*1501		GAPLGGAARALAHGVRVLED	Class-II	C L Day
			331			# *	GAPLGGAARALAHGVRVLED	Class-II	J Schulze zur Wiesch, 2012
			C70	DRB1*0101	DRB1*1501		GGAARALAHGVRVLE	Class-II	D Ciuffreda
			C77	DRB1*0101		# *	ALAHGVRVL	Class-II	H F Löhr
							LAHGVRVLEDGVNYATGNLP	Class-II	J Schulze zur Wiesch, 2012
									· ·
1423-1440	AYYRGLDVSVIPTAGDVV	NS3	C22	DRB1*0404	DRB1*1104		GINAVAYYRGLDVSVIPT <b>S</b> G	Class-II	J Schulze zur Wiesch, 2007
			S11	DRB1*0404	DRB1*0701	# *	GINAVAYYRGLDVSV	Class-II	A M Wertheimer
							GINAVAYYRGLDVSVIPT <b>S</b> G	Class-II	C L Day
							VAYYRGLDVSVIPT <b>S</b>	Class-II	A M Wertheimer
							LDVSVIPTSGDVVVVATDAL	Class-II	J Schulze zur Wiesch, 2007
							IPT SGDVVVVSTDALMTG	Class-II	N M Tabatabai
2548-2565	NQIRSVWEDLLEDTTTPI	NS5B	S2	DRB1*0701	DRB1*1454	# *	HARKAVTHINSVWKDLLEDN	Class-II	J Schulze zur Wiesch, 2012
							SVWKDLLED <b>NV</b> TPIDTTIMA	Class-II	C L Day
_				-			1		
2893-2908	IIERLHGLSAFTLHSY	NS5B	S2	DRB1*0701	DRB1*1454	# *	PIIQRLHGLSAFSLHSYSPG	Class-II	J Schulze zur Wiesch, 2007
				-			PIIQRLHGLSAFSLHSYSPG	Class-II	J Schulze zur Wiesch, 2012

 Table S8: Overlap between genotype-3-specific CD4+ T-cell targets defined in

 this study and previously described HCV genotype-1-specific CD4+ targets.

HCV genotype-3-specific CD4+ T-cell targets and described genotype-1-specific CD4+ targets and those without defined CD4/CD8 restricted were classified by overlap (left coloured bar): 'likely overlapping' (light blue, <20% sequence differences within targeted area), 'unlikely overlapping' (light pink, >20% differences within targeted area, or less than 7 amino acids overlap), and 'not overlapping' (pink). *Left:* For each epitope, viral region, position and genotype-3 sequence is given, as well as patients targeting the epitopes (colour coding: orange-SR, blue-acute, yellow chronic), and patient's HLA class-II types. *Right:* Published overlapping genotype-1 CD8+ epitopes are specified, including sequence, restricting HLA type and first author of the publication; overlap between epitopes is underlined and genotype-3/genotype-1 sequence differences marked bold. # CD4 restriction for marked patient experimentally defined; \* Targets previously described in (Humphreys et al. 2012). (Lasarte et al. 1998; Lamonaca et al. 1999; Löhr et al. 1999; Tabatabai et al. 1999; Day et al. 2002; MacDonald et al. 2002; Sugimoto et al. 2003; Wertheimer et al. 2003; Kennedy et al. 2006; Castelli et al. 2007; Schulze zur Wiesch et al. 2007; Ciuffreda et al. 2008; Schulze Zur Wiesch et al. 2012).



Figure S1: Comparison of breadth and targeted viral regions in spontaneously resolved individuals and patients with chronic HCV genotype-3 infection.

(A) Numbers of viral regions targeted in spontaneously resolved HCV infection and chronic HCV genotype-3 infection (unpaired t-test). (B) Comparison of numbers of patients targeting structural and non-structural regions in patient with chronic HCV genotype-3 infection and patients with spontaneously resolved HCV infection (Fisher's exact).





T-cell responses detected by IFNγ ELISpot assay to HLA predicted peptides and overlapping peptide pools (OP) were measured in patients with (A) acutely (n=16) and (B) chronically HCV genotype-3 infected (n=64) patients, and (C) spontaneously resolved infection (n=8). Responses to overlapping pools are compared to the HCV viral region in which the HLA predicted peptide falls (NS3p, NS3h, NS4, NS5a, NS5b1, NS5b2). SFU: Spot forming units.





### Figure S3: Evaluation of sequence heterogeneity at targeted HCV genotype-3-specific epitopes.

Sequence heterogeneity was evaluated by Shannon entropy scores in HCV viral regions (A) core, (B) E2, (C) NS2, (D) NS3, (E) NS4b, (F) NS5a and (G) NS5b on population level using Los Alamos genotype-3 sequences. The number of sequences used for each calculation is given. CD4+ (orange) and CD8+ (green) T-cell targets detected within the Oxford genotype-3 cohort are marked. (H) Mean Shannon entropy scores for CD4+ and CD8+ epitopes were calculated (p=0.3438, unpaired t-test). (I) Sequence polymorphisms within the Oxford cohort were evaluated at 16 CD8+ and 4 CD4+ T-cell targets in patients with detected T-cell responses. Sequence polymorphisms at CD4+ epitopes were detected in 2/19 sequenced patients, whereas polymorphisms at CD8+ epitopes were detected in 14/32 sequenced patients, respectively (p=0.0152, Fisher's exact).



Figure S4: T-cell responses detected against HCV genotype-1 peptide sets in spontaneously resolved patients and acutely infected patients with subsequent resolution of infection.

Hepatitis C virus (HCV) genotype-1 specific T-cell responses were measured by IFN $\gamma$ -ELISpot assays (spot-forming units (SFU)/10<sup>6</sup> peripheral blood mononuclear cells (PBMC)) using an HCV genotype-1b-specific peptide set spanning the entire HCV genome in 16 patients with spontaneously resolved infection (SR), and 4 patients acutely infected with HCV genotype-3 who subsequently spontaneously resolved infection (A-SR).



Figure S5: Hepatitis C virus (HCV) genotype-1 sequences variants at T-cell targets identified in spontaneously resolved patients.

HCV genotype-1 sequences were obtained from the Los Alamos database at T-cell targets identified in spontaneously resolved patients, to define common HCV genotype-1 sequence variants (frequencies >15%). For each T-cell target (A-R), HCV genotype-3 consensus sequence (top of each graph) and common HCV genotype-1 variants are specified. Where T-cell cross-reactivity to common variants has not been assessed experimentally due to lack of PBMC, sequences are marked with a star.

А

	Epitope s	equence	, Fé	Viral egion	start AA	end AA	HLA type	number of publications	T cell targets in Oxford gt3 cohort	Restriction of HCV gt3 epitopes
CD8	YLLPRRGE	RL	co	ore	35	44	A2	10		
	GPRLGFRA	Т	co	ore	41	49	B7	5		
	NEGLGWTG	W	co	ore	87	95	B44	5		
	ADLMGYIE	LV	co	ore	131	140	A2	11		
	LLALLSCI	TV	co	ore	178	187	A2	5		
	SMVGNWAR	V	E	1	363	371	A2	5		
	SLLAPGAR	ONV	E	2	401	411	A2	5		
	NTRPPLGN	~ W	E	2	541	549	B57	5		
	RDWAHNGI		N	52	957	964	B37	7		
	CINCUCWI	V	N	53	1073	1081	Δ2	31		
	LLODACUA	17	N	63	1160	1177	12	51		
	LLCPAGHA	.v	IN N	53	1169	11//	AZ	5		
	HPNIEEVA	L	N	53	1359	1367	B35	/		
	HSKKKCDE	L	N	53	1395	1403	88	8		
	KLVALGIN	AV	N	S3	1406	1415	A2	32		
	ATDALMIG	Y	N	S3	1436	1444	A1	7	+	CD8+
	LLFNILGO	WV	N	S4b	1807	1816	A2	7		
	VLSDFKTW	L	N	S4a	1987	1995	A2	7		
	ALYDVVTR	T.	N	S5b	2594	2602	A2	11		
		-								
CD4+	NKRNTNRF	PQDVKFPGGGQIVGGVYLLPRF	GPRL CO	ore	11	44	ND	9		65.4
	KVRRPEGF	TWAQPGYPWPLYGNEGLGWAGW	LLSPRGS CC	ore	6/	103	ND	10	+	CD4+
	ADLMGYIF	LVGAPLGGAARALAHGVRVLEI	co	ore	131	160	ND	12	+	CD4+
	ATRDGKLE	ATQLRRHIDLL	E	1	247	265	ND	7		
	YFSMVGNW	AKVLVVL	E	1	361	375	ND	6		
	SSDLYLVI	RHADVIP	N	S3	1127	1141	ND	6		
	LETTMRSF	VFTDNSSPPVVP	N	S3	1201	1220	ND	6		
	PAAYAAOG	YKVLVLNPSVAA	N	S3	1241	1260	ND	8		
	LADAGOSO	GAYDITICDECHSTDAT	N	53	1300	1324	ND	8		
	FULLCODE	I TECHEVEVED	N	C3	1202	1401	ND	0		
	CNIECUIRO	NDEGL DDEE	IN N	55	1454	1401	ND	7		
	CNICVIQI	VDFSLDPIFI	IN .	55	1434	14/1	ND	/		
	FVAPGERE	SGMFD	N	\$3	1501	1513	ND	6		
	YELTPAET	TVRLRAYMNTPGLPVAQD	N	S3	1528	1553	ND	11		
	THIDAHFI	SQTK	N	S3	1566	1577	ND	6		
	ENLPYLVA	YQATVCARAQAPPPSW	N	S3	1581	1604	ND	10		
	PLLYRLGA	VQNEITLTHP	N	S3	1623	1640	ND	8		
	VIVGRVVI	SGKPAIIPDREV	N	S4a	1681	1700	ND	6		
	GLSTLPGN	PAIASL	N	S4a	1777	1790	ND	6		
	ALWVGVVC	AATLERHVGPGE	N	S4a	1891	1910	ND	6		
	OCCDUDRO	ADVATKSLTEDI	N	S5h	2661	2680	ND	6		
	KLVALGINA	V-				1	CINGV	CWTV-		
	V						-V			
	M									
		2						n a		
	M-V					\$2	A	<b>b</b>		
	G					tian				
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ark	GV					È		SI-		
Ê	SGL					-	-VG			
G	66 T							E C		
							-VA	·		
	SL						-V	s-		
	Q-SG	-			iants		-L			
		- <b>L</b>			Var			ſ		
						·	-VS			_
	sv						TVG	M	_	
	TGL						TIG	M	-	
\$	RGM-L	-				ants	T-G	M		
riant.	R-RGM-L					varić	TVG	I		
3 (a)	RGL-L					313	TIC	M		
GT	I-RGM-T					Ĭ	116			
	PCM V		6T1a GT1b				SVG	M		
	RGM-V		GT3a			1	AVG	M		G G
	other	S-					ot	hers		G
		0 20 40 60	80 100					0 20	40 6	30 80
		% conuencee								

# Figure S6: T-cell targets frequently detected in HCV genotype-1 infection are not targeted in HCV genotype-3 infection, likely due to sequence differences between genotypes.

(A) Comparison of CD8+ restricted epitopes dominant in HCV genotype-1 infection (defined as described in 5 or more publications on the IEDB). If the epitope was targeted in the Oxford HCV genotype-3 cohort, it is marked with +. Epitopes described as CD4+ restricted epitopes in the Oxford genotype-3 cohort falling into regions of CD8+ restricted epitopes described as HCV genotype-1 epitopes in the literature are marked in grey. (B) Comparison of circulating viral genotype-1 and genotype-3 sequences at dominant HCV genotype-1 epitopes. Sequences were obtained from the Los Alamos database, with additional in-house genotype-3 sequences.

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